PSEUDOCYPHELLARINS A AND B, TWO FULLY SUBSTITUTED DEPSIDES FROM THE LICHEN PSEUDOCYPHELLARIA ENDOCHRYSEA*†

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Key Word Index—Pseudocyphellaria endochrysea; Stictaceae; lichen; pseudocyphellarin A; pseudocyphellarin B; depsides.

Abstract—The structures of two fully substituted depsides, pseudocyphellarins A and B, have been elucidated by spectroscopic and chemical methods from the lichen Pseudocyphellaria endochrysea.

INTRODUCTION

In the course of our chemical investigations of Antarctic lichens [1] we isolated from *Pseudocyphellaria endochrysea* two new compunds, pseudocyphellarin A and pseudocyphellarin B. The structural elucidation of both substances is described in the present paper.

RESULTS AND DISCUSSION

Pseudocyphellarin A, mp 173–175°, has according to the high resolution mass spectrum the formula $C_{21}H_{22}O_8$ (found m/z 402.1306; calculated 402.1315) and a UV spectrum with $\lambda_{\max}^{\text{MeOH}}$ (log ε) 218 (4.15), 250 (4.36), S 270 (4.21), S 290 (3.96) and 342 nm (3.30) and $\lambda_{\max}^{\text{MeOH}+\text{NaOH}}$ (log ε) 221 (4.29), 286 (4.19) and S 302 (4.13) similar to the UV spectrum of nephroarctin with $\lambda_{\max}^{\text{MeOH}}$ (log ε) 238 (3.81), S 259 (3.97), 281 (4.09), 315 (4.00) and 379 nm (3.54) [2]. Pseudocyphellarin A gave a yellow colour with p-phenylenediamine, indicative of an aldehyde group. The ¹H NMR spectrum (100MHz, CDCl₃) of pseudocyphellarin A showed five methyl signals at δ 2.05, 2.06, 2.15, 2.45 and 2.70, a methoxy group signal at δ 3.95, an aldehyde signal at δ 10.34 and three hydroxyl group signals at δ 11.06, 12.32 (br) and 13.00, proving the presence of a fully substituted depside.

To discover the disribution of the subsituents at the Sand A-parts of the depside, pseudocyphellarin A was submitted to tert.-butanolysis [3,4], which gave after chromatography two main products (S and A) and a minor one (S'). Compound S, mp 117–119°, gave in the mass spectrum a [M]⁺ peak at m/z 266 corresponding to the formula $C_{14}H_{18}O_5$, in the ¹H NMR spectrum (100 MHz, CDCl₃) signals at δ 1.60 (s, 9H, -CMe₃), 2.07 (s, 3H, -Me), 2.44 (s, 3H, -Me), 10.24 (s, 1H, -CHO), 12.44 (br s, 1H, -OH) and 12.70 (s, 1H, -OH) and proved to be tert.-butyl 2,4-dihydroxy-3-formyl-5,6-dimethylbenzoate (1). Saponification of 1 with KOH-MeOH led to 2,4-dihydroxy-3-formyl-5,6-dimethylbenzene (2) [5]. The minor cleavage product S' was identical with 2. Compound A, mp 92-94°, showed in the ¹H NMR spectrum (100 MHz, CDCl₃) signals at δ 2.11 (s, 6H, 2 ×-Me), 2.40 (s, 3H, -Me), 3.91 (s, 3H, -CO₂Me), 5.90 (br s, 1H, -OH) and 11.38 (s, 1H, -OH) and proved to be identical with an authentic sample of methyl 2,4-dihydroxy-3,5,6-trimethylbenzoate (3) (IR, mmp and R_f in three different solvent systems) [6]. Cleavage product 3 gave on hydrolysis with KOH-H₂O 2,4-dihydroxy-3,5,6-trimethylbenzene (4).

Hence the S- and A-parts of pseudocyphellarin A are 2,4-dihydroxy-3-formyl-5,6-dimethylbenzoic acid and 2,4-dihydroxy-3,5,6-trimethylbenzoic acid, respectively. Because in nearly all of the naturally occurring depsides the p-hydroxyl group of the A-part of the molecule is connected to the S-part, pseudocyphellarin A should have structure 5. This structure was finally proved by Dr. J. A. Elix (personal communication) who synthesized compound 5 and found it to be identical with pseudocyphellarin A in all respects (¹H NMR, mp, mmp and TLC R_c values in several solvents). Acetylation of 5 with Ac₂O-H₂SO₄ gave the pentaacetate 6 and methylation with dimethyl sulphate-potassium carbonate in dimethyl formamide yielded the tri-O-methyl ether 7. Alkaline hydrolysis of 7 gave 2,4-di-O-methyl-3-formyl-5,6-dimethylbenzoic acid (8). Hydrogenation of 5 with palladium on charcoal led to hypopseudocyphellarin A

Pseudocyphellarin B, mp $168-169^{\circ}$, showed in the ¹H NMR spectrum (100 MHz, CDCl₃-DMSO- d_6) five methyl signals at δ 2.02 (s, 6H, 2 × -Me), 2.11, 2.35 and 2.57 (3 × s, 3 × 3H, 3 × -Me), a methoxy group signal at δ 3.92, a signal of a benzylic hydroxyl group at δ 4.89 (s, 2H, -CH₂OH) and three hydroxyl group signals at δ 3.13 (br s, 1H, -OH), 10.33 (br s, 2H, 2 × -OH) and 11.33 (s, 1H, -OH). The mass spectrum of pseudocyphellarin B did not show a [M]⁺ but gave peaks at m/z 210 and 178, corresponding to the fragment ions a, $C_{11}H_{14}O_4$ (found 210.0896; calculated 210.0892), and b, $C_{10}H_{10}O_3$ (found 178.0632; calculated 178.0630). On acetylation of

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pseudocyphellarin B with $Ac_2O-H_2SO_4$ a tetraacetate, mp 145–146°, resulted, the ¹H NMR spectrum (100 MHz, CDCl₃) of which showed the following signals: δ 1.97, 1.99, 2.08, 2.12, 2.17 (5 × s, 5 × 3H, 5 × -Me), 2.23 (br s, 6H, 2 × -Me), 2.32 and 2.44 (2 × s, 2 × 3H, 2 × -Me), 3.85 (s, 3H, -CO₂Me) and 4.96 (s, 2H, -CH₂-OAc). tert.-Butanolysis of pseudocyphellarin B gave surprisingly 2,6-dihydroxy-3,4-dimethyl-5-tert.-butoxycarbonyl-1-benzyl tert.-butyl ether (10) and methyl 2,4-dihydroxy-3,5,6-trimethylbenzoate (3), identical in all respects with the cleavage product of pseudocyphellarin A. These results and biogenetic considerations led to structures 11 and 12 of pseudocyphellarin B and its tetraacetate, respectively.

The comparison of the chemical shifts of the methyl group signals of pseudocyphellarin A (5), pentaacetylpseudocyphellarin A (6), tri-O-methylpseudocyphellarin A (7), hypopseudocyphellarin A (9), pseudocyphellarin B (11), tetraacetylpseudocyphellarin B (12), nephroarctin (13), methyl 2,4-dihydroxy-3,5,6-trimethylbenzoate (3), 2,4-dihydroxy-3,5,6-trimethylbenzene (4) and methyl 2,4dihydroxy-3,6-dimethylbenzoate (14) (Table 1) reveals an interesting phenomenon. Only in depsides with a free 2hydroxyl group (5, 9, 11 and 13) do the chemical shifts of the methyl groups at C-5' appear between δ 2.57 and 2.73. What is the reason for these high values? One can assume that there is a strong hydrogen bridge between the hydroxyl proton at C-2 and the depside carbonyl group. This is proved by the IR spectra of 5, 9, 11 and 13 with carbonyl bands at 1630 and 1640 cm⁻¹. Assuming a more or less planar conformation of the whole molecule, the methyl group either at C-3' or C-5' comes very close to the depside carbonyl group and is thus deshielded. In the depsides with an acetoxy or methoxy group at C-2 no hydrogen bond to the depside carbonyl group is possible and in consequence the molecule has another non-planar conformation.

The most important fragment ions of the mass spectra of 1, 3, 5 and 11 are shown in Scheme 1.

Besides pseudocyphellarins A and B, the following compounds were isolated from *P. endochrysea*: 3-oxostictan-22 α -ol, 2 α -acetoxystictan-3 β ,22 α -diol, stictan-2 α ,3 β ,22 α -triol, a mixture of polyols and a mixture of triacylglycerols with palmitic, oleic, linoleic and linolenic acids as components.

Scheme 1. Mass spectral fragmentations of compounds 1, 3, 5 and 11.

EXPERIMENTAL

Extraction. Air-dried and pulverized *P. endochrysea* (Del.) Vain. (198.5 g; from South Georgia, Dartmouth Point, leg. et det. R. I. Lewis Smith, 2 March 1982; voucher specimen deposited at the herbarium of S.H.) was extracted with Et_2O for 20 hr, the extract freed from solvent, the residue dissolved in CHCl₃, adsorbed on silica gel (with 5% H₂O; 50 g) and put on the top of a column with silica gel (with 5% H₂O; 500 g) in *n*-hexane. Elution with *n*-hexane- Et_2O (2 l., 9:1) gave a small amount of wax which was not investigated further. *n*-Hexane- Et_2O (1 l., 425:7.5) eluted pseudocyphellarin A (0.8 g, 0.4%), *n*-hexane- Et_2O (2 l., 4:1) an oily mixture of triacylglycerols (0.73 g, 0.36%) and *n*-hexane- Et_2O (1 l., 0.7:0.3) pseudocyphellarin B. Further elution of the column with *n*-hexane- Et_2O (2 l., 1:1) gave 3-oxostictan-22α-ol, mp 213-215° (0.4 g; 0.2%) and 2α-acetoxystictan-3β,22-diol, mp 214-216° and $[\alpha]_{24}^{24} - 33$ ° (CHCl₃; *c* 0.75) (1.33 g; 0.67%). Elution of the column with Et_2O -MeOH (500 ml, 9:1)

Table 1. ¹H NMR chemical shifts of the methyl groups in compounds 3-7, 9 and 11-14

Compound	Chemical shift of the methyl group at		
	C-3'	C-5'	C-6' respectively
	C-3	C-5	C-6
Pseudocyphellarin A (5)	2.45	2.70	2.15
Pentaacetylpseudocyphellarin A (6)	2.45	2.36	2.14
Tri-O-methylpseudocyphellarin A (7)	2.40	2.22	2.19
Hypopseudocyphellarin A (9)	2.40	2.65	2.29
Pseudocyphellarin B (11)	2.35	2.57	2.11
Tetraacetylpseudocyphellarin B (12)	2.44	2.32	2.12 or 2.17
Nephroarctin (13)	2.27	2.73	2.10
Methyl 2,4-dihydroxy-3,5,6-			
trimethylbenzoate (3)	2.40	2.11	2.11
2,4-Dihydroxy-3,5,6-trimethyl-			
benzene (4)	2.15	2.10	2.07
2,4-Dihydroxy-3,6-dimethyl-			
benzene (14)	2.30		2.00

5 R=H, R'=CH0
6 R=Ac, R'=CH(OAc)₂
7 R=Me, R'=CH0
9 R=H, R'=Me
11 R=H, R'=CH₂OH

gave stictan-2 α ,3 β ,22 α -triol, mp 256–257° and $[\alpha]_D^{24} + 6^\circ$ (CHCl₃; c 0.3) (50 mg; 0.02 %).

Extraction of the lichen with Me₂CO gave after recrystallization from EtOH a mixture of polyols as needles, mp 148-152° (90 mg; 0.04%).

Pseudocyphellarin A (5). Prisms, mp 173–175° (from Me₂CO) and the following colour reactions: KOH yellow, p-phenylene-diamine yellow and FeCl₃ (in EtOH) red-brown. $R_f = 0.67$ (silica gel Merck PF 254 + 366, n-hexane–Et₂O–HCO₂H, 30: 20: 6, grey spot after heating with SO₃HCl–HOAc). C₂₁H₂₂O₈ (402.4). IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 740, 782, 810, 852, 890, 926, 942, 1010, 1030, 1070, 1092, 1116, 1190, 1290, 1312, 1428, 1640, 2980, 3400. MS m/z (rel. int.): 402 [M]⁺ (6), 210.0896 [a]⁺ (97); calc. for C₁₁H₁₄O₄: 210.0892, 178.0633 [b]⁺ (100); calc. for C₁₀H₁₀O₃: 178.0630,

164.0474 [c]⁺ (50); calc. for $C_9H_8O_3$: 164.0473, 150.0684 [d]⁺ (92); calc. for $C_9H_{10}O_2$: 150.0682, 136.0522 [e]⁺ (16); calc. for $C_8H_8O_2$: 136.0524. ¹³C NMR (50.32 MHz, CDCl₃): C-1: δ 102.9 (s), C-2: 167.0 (s), C-3: 108.0 (s), C-4: 166.1, C-5: 118.2 (s), C-6: 151.5 (s), C-7: 169.7 (s), C-8: 18.8 (q), C-9: 10.7 (q), C-10: 194.0 (d), C-1': 116.2 (s), C-2': 159.0 (s), C-3': 111.9 (s), C-4': 150.1 (s), C-5': 120.5, C-6': 137.6 (s), C-7': 172.1 (s), C-8': 20.4 (q), C-9': 13.2 (q), C-10': 9.7 (q), C-11': 52.3 (q).

Pseudocyphellarin B (11). Needles, mp $168-169^{\circ}$ (dec.) and a blue-violet colour reaction with FeCl₃ (in EtOH). $C_{21}H_{24}O_{8}$ (404.4). UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 226 (4.30), 275 (4.25), 318 (3.93); $\lambda_{\max}^{\text{MeOH}+\text{NaOH}}$ nm (log ε): 228 (4.30), 245 (4.20), 322 (4.41). IR ν_{\max}^{KBr} cm⁻¹: 740, 776, 804, 890, 966, 990, 1002, 1070, 1092, 1110, 1170, 1240, 1260, 1318, 1440, 1578, 1604, 1640, 2960, 3250, 3550. MS, m/z (rel. int.): 210.0896 [a] + (83); calc. for $C_{11}H_{14}O_4$: 210.0892, 178.0632 [b] + (92); calc. for $C_{10}H_{10}O_3$: 178.0630, 150.0691 [d] + (100); calc. for $C_{9}H_{10}O_2$: 150.0692.

tert.-Butanolysis of 5. 5 (0.18 g) in tert.-BuoH was heated under reflux for 48 hr. After this time TLC showed the presence of three products with R_f values 0.88 (A), 0.60 (B) and 0.52 (C), A and B being the main products. The residue after removal of solvent was dissolved in C₆H₆ and chromatographed on silica gel (7 g, with 5% H_2O); C_6H_6 (100 ml) eluted tert.-butyl 2,4dihydroxy-3-formyl-5,6-dimethylbenzoate (A, 1) as needles, mp 117-119° (from n-hexane), and the following colour reactions: KOH yellow to yellow-orange, p-phenylenediamine yellow, FeCl₃ (in EtOH) dirty green, on dilution with H₂O dirty blue, KOH + NaOCl orange-red. $C_{14}H_{18}O_5$ (266.3). UV λ_{max}^{MeOH} nm (log ε): 243 (4.63), 269 (4.56), S 290 (4.40); λ_{max}^{MeOH} nm $(\log \varepsilon)$: 220 (4.54), 292 (4.47), 410 (4.04). MS m/z (rel. int.): 266 $[M]^+$ (30), 210 $[f]^+$ (97), 192 $[g]^+$ (83), 164 $[c]^+$ (100), 136 $[e]^+$ (53), 108 [h] + (24). Further elution of the column with C₆H₆ (100 ml) gave methyl 2,4-dihydroxy-3,5,6-trimethylbenzoate (B, 3) as needles, mp 92-94° (from n-hexane), and the following colour reactions: NaOCl deep red, FeCl₃ (in EtOH) blue. $C_{11}H_{14}O_4$ (210.2). UV λ_{max}^{McOH} nm (log ε): 226 (4.71), 270 (4.66), 314 (4.24); $\lambda_{max}^{McOH+NaOH}$ nm (log ε): 221 (4.66), 246 (4.58), 317 (4.89). MS m/z (rel. int.): 210 [M]⁺ (91), 178 [b]⁺ (100), 150 [d]⁺ (98), 122 [i] + (63). Finally C₆H₆ (300 ml) eluted 2,4-dihydroxy-3formyl-5,6-dimethylbenzene (C, 2) as yellow needles, mp 137–139° (from *n*-hexane). $C_9H_{10}O_3$ (166.2). IR v_{max}^{KBr} cm⁻¹: 702, 730, 756, 840, 892, 1000, 1100, 1204, 1242, 1290, 1340, 1380, 1416, 1444, 1510, 1598, 1626, 2920, 3200. ¹H NMR (200 MHz, CDCl₃): δ 2.04 (s, 3H, C-5-Me), 2.22 (s, 3H, C-6-Me), 6.14 (s, 1H, aromatic-H), 10.27 (s, 1H, -CHO). MS m/z (rel. int.): 166 [M]⁺ (100), 151 $[M - Me]^+$ (72), 137 $[M - CO - H]^+$ (68).

Hydrolysis of tert.-butyl 2,4-dihydroxy-3-formyl-5,6-dimethylbenzoate. 1, (0.12 g) was heated with KOH (0.3 g) in $\rm H_2O$ (2.5 ml) under $\rm H_2$ under reflux for 8 hr. After usual work-up and chromatography on silica gel (6 g, with 5 % $\rm H_2O$), $\rm C_6H_6$ (500 ml) eluted 2,4-dihydroxy-3-formyl-5,6-dimethylbenzene (2) as yellow needles, mp 137–139° (from n-hexane), identical with compound C from the tert.-butanolysis of pseudocyphellarin A.

The 2,4-dinitrophenylhydrazone of 2 was orange-red needles, mp 285–287° (dec., from EtOH–EtOAc). Robertson and Whalley [5] reported mp 140° for the aldehyde 2 and mp 289° for the corresponding 2,4-dinitrophenylhydrazone. IR $v_{\rm max}^{\rm KBr}$ cm⁻¹: 710, 744, 820, 840, 912, 930, 970, 1004, 1080, 1130, 1144, 1220, 1240, 1270, 1304, 1330, 1380, 1422, 1460, 1500, 1518, 1610, 2980, 3150, 3350, 3500.

Hydrolysis of methyl 2,4-dihydroxy-3,5,6-trimethylbenzoate. 3 (0.1 g) was heated with KOH (0.6 g) in H_2O (5 ml) under H_2 under reflux for 2 hr. After usual work-up and chromatography on silica gel (5 g, with 5 % H_2O), C_6H_6 (100 ml) eluted 2,4-dihydroxy-3,5,6-trimethylbenzene (4), silk-like needles, mp 145-146° (from Et_2O -n-hexane). $C_9H_{12}O_2$ (152.2).

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IR $v_{\rm max}^{\rm KBr}$ cm $^{-1}$: 840, 922, 1002, 1030, 1078, 1160, 1200, 1240, 1290, 1318, 1380, 1468, 1510, 1590, 1622, 2980, 3300, 3500. 1 H NMR (100 MHz, CDCl₃): δ 2.07, 2.10, 2.15 (3 × s, 3 × 3H, 3 × -Me), 5.87, 6.25 (2 × s, 2 × 1H, 2 × -OH), 7.30 (s, 1H, aromatic -H).

Pentaacetylpseudocyphellarin A (6). From 5 (0.1 g) and $Ac_2O-H_2SO_4$ (2 ml of a mixture of 5 ml Ac_2O and 1 drop of conc. H_2SO_4) at room temp. in 24 hr. After usual work-up and crystallization from CHCl₃–MeOH, prismatic plates, mp 198–200°. $C_{31}H_{32}O_{14}$ (628.6). IR v_{max}^{KBr} cm⁻¹: 790, 870, 900, 918, 1008, 1070, 1080, 1104, 1150, 1200, 1228, 1270, 1322, 1370, 1440, 1580, 1602, 1760, 2990, 3520. ¹H NMR (100 MHz, CDCl₃): δ 2.02 (s, 9H, 3 × -Me), 2.08, 2.14 (2 × s, 2 × 3H, 2 × -Me), 2.23 (s, 9H, 3 × -Me), 2.36, 2.45 (2 × s, 2 × 3H, 2 × -Me), 3.84 (s, 3H, -CO₂Me)

Tri-O-methylpseudocyphellarin A (7). To 5 (0.13 g) and $\rm K_2CO_3$ (0.5 g) in DMF (3 ml) was added Me₂SO₄ (0.25 ml) and the mixture heated at 100° for 10 min. $\rm K_2CO_3$ (0.3 g) and Me₂SO₄ (0.2 ml) were then added and heating was continued for 15 min. After dilution with H₂O, the ppt. was collected by filtration, dried at room temp. and chromatographed in C₆H₆ on silica gel (5 g, with 5% H₂O); C₆H₆ (400 ml) eluted the tri-O-methyl ether, prisms, mp 150–152° (from CHCl₃–n-hexane). C₂₄H₂₈O₈ (444.5). IR $\rm v_{max}^{KBr}$ cm⁻¹: 782, 830, 870, 910, 936, 958, 984, 1010, 1030, 1072, 1100, 1154, 1204, 1280, 1292, 1368, 1332, 1390, 1466, 1578, 1684, 1730, 3000. ¹H NMR (100 MHz, CDCl₃): δ 2.19 (s, 6H, 2 × -Me), 2.22 (s, 6H, 2 × -Me), 2.40 (s, 3H, -Me), 3.75, 3.84 (2 × s, 2 × 3H, 2 × -OMe), 3.91 (s, 6H, -OMe, -CO₂Me), 10.32 (s, 1H, -CHO).

2,4-Dimethoxy-3-formyl-5,6-dimethylbenzoic acid (8). By saponification of 7 (20 mg) with KOH (0.5 g) in MeOH (5 ml) under reflux for 2 hr. The reaction mixture was acidified with $10\,\%$ H₂SO₄, extracted with Et₂O, the Et₂O extract shaken with NaHCO₃ soln (5%), the NaHCO₃ extract acidified and again extracted with Et₂O. The Et₂O was removed and the residue twice recrystallized from MeOH-H₂O: prisms, mp $112-115^\circ$. C₁₂H₁₄O₅ (238.2). IR $\nu_{\rm max}^{\rm KBr}$ cm⁻¹: 760, 902, 952, 984, 1010, 1046, 1090, 1110, 1190, 1290, 1380, 1410, 1464, 1580, 1682, 1722, 3000, 3200. MS m/z (rel. int.): 238 [M]⁺ (100), 220 [M - H₂O]⁺ (67), 205 [M - H₂O - Me]⁺ (53), 177 [M - H₂O - Me - CO]⁺ (40).

Hypopseudocyphellarin A (9). By hydrogenation of 5 (0.1 g) in EtOAc (30 ml) with 10 % Pd/C (0.1 g) under normal conditions for 6 hr and chromatography of the resulting product on silica gel (6 g, with 5 % H₂O). After elution with C₆H₆ (200 ml), Et₂O (200 ml) eluted hypopseudocyphellarin A, rectangular prisms, mp 172–173° (dec., from CHCl₃–MeOH). C₂₁H₂₄O₇ (388.4). IR $\nu_{\rm max}^{\rm KB}$ cm⁻¹: 746, 780, 810, 900, 970, 990, 1010, 1030, 1070, 1100, 1114, 1170, 1240, 1262, 1310, 1398, 1444, 1580, 1610, 1640, 3020, 3250, 3550. ¹H NMR (100 MHz, DMSO-d₆): δ 2.29 (s, 6H, 2 × –Me), 2.34, 2.40, 2.65, 3.59 (4 × s, 4 × 3H, 4 × –Me), 4.10 (s, 3H, –CO₂Me), 9.40, 9.97, 10.59 (3 × s, 3 × H, 3 × –OH).

tert.-Butanolysis of pseudocyphellarin B. 11 (50 mg) in tert. BuOH (50 ml) was heated under reflux for 24 hr. The residue after removal of the solvent was separated by prep. TLC. (silica gel, Merck PF 254+366, $10 \times 10 \times 0.1$ cm, n-hexane-Et₂O-HCO₂H, 30:20:3) and showed two main bands of R_f values 0.56 (A) and 0.80 (B). Elution of band A gave 2,6-dihydroxy-3,4-dimethyl-5-tert.-butoxycarbonyl-1-benzyl-tert.

butyl ether (10), needles, mp 69–70° (from MeOH–H₂O). $C_{18}H_{28}O_5$ (324.4). IR v_{max}^{KBr} cm⁻¹: 762, 784, 844, 878, 894, 1006, 1050, 1084, 1116, 1150, 1220, 1260, 1286, 1336, 1360, 1394, 1464, 1570, 1624, 3000, 3220, 3500. ¹H NMR (100 MHz, CDCl₃): δ 1.28, 1.56 (2 × s, 2 × 9H, 2 × -CMe₃), 2.03, 2.36 (2 × s, 2 × 3H, 2 × -Me), 4.78 (s, 2H, -CH₂–O-), 9.65, 11.61 (2 × s, 2 × H, 2 × -OH). MS m/z (rel. int.): 324 [M]⁺ (75), 268 [M - CH₂ = CMe₂]⁺ (77), 250 [M - CH₂ = CMe₂ - H₂O]⁺ (62), 232 [M - CH₂ = CMe₂ - 2H₂O]⁺ (45), 194 [M - 2CH₂ = CMe₂ - H₂O]⁺ (98), 166 [M - 2CH₂ = CMe₂ - H₂O]⁺ (75). Elution of band B gave needles, mp 92–94° (from n-hexane), identical with 2,4-dihydroxy-3,5,6-trimethylbenzoate (3) in all respects.

Tetraacetylpseudocyphellarin B (12). From 11 (35 mg) and $Ac_2O-H_2SO_4$ (1 ml of a mixture of 5 ml Ac_2O and 1 drop of conc. H_2SO_4) at room temp. in 24 hr. After usual work-up and crystallization from MeOH, small prisms, mp 145–146°. $C_{29}H_{32}O_{12}$ (572.5). IR v_{max}^{KBr} cm⁻¹: 796, 880, 920, 1030, 1074, 1150, 1180, 1206, 1244, 1268, 1324, 1372, 1440, 1574, 1608, 1730, 1762, 2980

GC/MS analysis of triacylglycerols. Part of the mixture of triacylglycerols from P. endochrysea (0.1 g) was hydrolysed with KOH-MeOH and the resulting mixture of fatty acids converted into the corresponding methyl esters with CH₂N₂. GC/MS (10% EGSS-X on Gaschrom P, 125–150 μ m, glass column, 2 mm i.d., 180 cm, $T=160^{\circ}$) showed the presence of methyl palmitate (24%, m/z 270, C₁₇H₃₄O₂), methyl stearate (3%, m/z 298, C₁₉H₃₈O₂), methyl oleate (31%, m/z 296, C₁₉H₃₆O₂), methyl linoleate (35%, m/z 294, C₁₉H₃₄O₂) and methyl linolenate (5%, m/z 292, C₁₉H₃₂O₂).

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